INTENDED USE

The Alpha 2-Macroglobulin test kit is a highly sensitive two-site enzyme linked immunoassay (ELISA) for measuring Alpha 2-Macroglobulin in the serum and plasma of humans.

INTRODUCTION

Alpha 2-Macroglobulin (A2M) is a major protease inhibitor in serum and an acute phase protein which increases significantly in concentration in the human as a result of inflammation. The major pathophysiological role for human alpha 2-macroglobulin has yet to be conclusively defined. This kit is specific to the alpha 2-macroglobulin and will not cross react with the closely related alpha 1-macroglobulin.

PRINCIPLE OF THE ASSAY

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the Alpha 2-Macroglobulin present in samples reacts with the anti-Alpha 2-Macroglobulin antibodies which have been adsorbed to the surface of polystyrene microtitre wells. After the removal of unbound proteins by washing, anti-A2M antibodies conjugated with horseradish peroxidase (HRP), are added. These enzyme-labeled antibodies form complexes with the previously bound A2M. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme varies directly with the concentration of A2M in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of A2M in the test sample. The quantity of A2M in the test sample can be interpolated from the standard curve constructed from the standards, and corrected for sample dilution.

REAGENTS (Quantities sufficient for 96 determinations)

1. DILUENT CONCENTRATE (Running Buffer)
   One bottle containing 50 ml of a 5X concentrated diluent running buffer.

2. WASH SOLUTION CONCENTRATE
   One bottle containing 50 ml of a 20X concentrated wash solution.

3. ENZYME-ANTIBODY CONJUGATE 100X
   One vial containing 150 μL of affinity purified anti-Human Alpha 2-Macroglobulin antibody conjugated with horseradish peroxidase in a stabilizing buffer.

4. CHROMOGEN-SUBSTRATE SOLUTION
   One vial containing 12 mL of 3,3',5,5'-tetramethylbenzidine (TMB) and hydrogen peroxide in citric acid buffer at pH 3.3.

5. STOP SOLUTION
   One vial containing 12 ml 0.3 M sulfuric acid.

WARNING: Avoid contact with skin.
6. ANTI-HUMAN A2M ELISA MICRO PLATE
Twelve removable eight (8) well micro well strips in well holder frame. Each well is coated with affinity purified anti-Human A2M.

7. HUMAN A2M CALIBRATOR
One vial containing a lyophilized Human Alpha 2-Macroglobulin calibrator.

FOR IN VITRO USE ONLY

REAGENT PREPARATION

1. DILUENT CONCENTRATE
The Diluent Solution supplied is a 5X Concentrate and must be diluted 1/5 with distilled or deionized water (1 part buffer concentrate, 4 parts dH2O).

2. WASH SOLUTION CONCENTRATE
The Wash Solution supplied is a 20X Concentrate and must be diluted 1/20 with distilled or deionized water (1 part buffer concentrate, 19 parts dH2O). Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35°C before dilution can dissolve crystals.

3. ENZYME-ANTIBODY CONJUGATE
Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 µL Enzyme-Antibody Conjugate to 990 µL of 1X Diluent for each test strip to be used for testing. Mix uniformly, but gently. Avoid foaming.

4. CHROMOGEN-SUBSTRATE SOLUTION
Ready to use as supplied.

5. STOP SOLUTION
Ready to use as supplied.

6. ANTI-HUMAN A2M ELISA MICRO PLATE
Ready to use as supplied. Unseal Microtiter Pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal along with desiccant.

7. HUMAN A2M CALIBRATOR
Add 1.0 ml of distilled or de-ionized water to the Human Alpha 2 Macroglobulin Calibrator and mix gently until dissolved. The calibrator is now at a concentration of 105.73 µg/ml (the reconstituted calibrator should be aliquoted and frozen if future use is intended). Human Alpha 2 Macroglobulin standards need to be prepared immediately prior to use (see the following chart). Mix well between each step. Avoid foaming.

<table>
<thead>
<tr>
<th>Standard</th>
<th>ng/ml</th>
<th>Volume added to 1x Diluent</th>
<th>Volume of 1x Diluent</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td>640</td>
<td>5 µL Human A2M Calibrator</td>
<td>821 µL</td>
</tr>
<tr>
<td>5</td>
<td>320</td>
<td>300 µL standard 6</td>
<td>300 µL</td>
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<tr>
<td>4</td>
<td>160</td>
<td>300 µL standard 5</td>
<td>300 µL</td>
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<tr>
<td>3</td>
<td>80</td>
<td>300 µL standard 4</td>
<td>300 µL</td>
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<tr>
<td>2</td>
<td>40</td>
<td>300 µL standard 3</td>
<td>300 µL</td>
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<tr>
<td>1</td>
<td>20</td>
<td>300 µL standard 2</td>
<td>300 µL</td>
</tr>
<tr>
<td>0</td>
<td>0</td>
<td></td>
<td>600 µL</td>
</tr>
</tbody>
</table>

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.
INDICATIONS OF INSTABILITY
If the test is performing correctly, the results observed with the standard solutions should be within 20 % of the expected values.

SPECIMEN COLLECTION AND HANDLING
Blood should be collected by venipuncture. The serum should be separated from the cells after clot formation by centrifugation. For plasma samples, blood should be collected into a container with an anticoagulant and then centrifuged. Care should be taken to minimize hemolysis, excessive hemolysis can impact your results. Assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles.

1. Precautions
For any sample that might contain pathogens, care must be taken to prevent contact with open wounds.

2. Additives and Preservatives
No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

3. Known interfering substances
Azide and thimerosal at concentrations higher than 0.1% inhibits the enzyme reaction.

MATERIAL PROVIDED
See "REAGENTS"

MATERIALS REQUIRED BUT NOT PROVIDED
• Precision pipette (2 μL to 200 μL) for making and dispensing dilutions
• Test tubes
• Microtitre washer/aspirator
• Distilled or Deionized H₂O
• Microtitre Plate reader
• Assorted glassware for the preparation of reagents and buffer solutions
• Timer

ASSAY PROTOCOL
DILUTION OF SAMPLES
The assay for quantification of A2M in samples requires that each test sample be diluted before use. For a single step determination a dilution of 1/2,000 is appropriate for most serum/plasma samples. For absolute quantification, samples that yield results outside the range of the standard curve, a lesser or greater dilution might be required. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

1. To prepare a 1/2,000 dilution of sample, transfer 5 μL of sample to 495μL of 1X diluent. This gives you a 1/100 dilution. Next, dilute the 1/100 samples by transferring 20 μL, to 380 μL of 1X diluent. You now have a 1/2,000 dilution of your sample. Mix thoroughly at each stage.

PROCEDURE
1. Bring all reagents to room temperature before use.

2. Pipette 100 μL of
   Standard 0 (0.0 ng/ml) in duplicate
   Standard 1 (20 ng/ml) in duplicate
   Standard 2 (40 ng/ml) in duplicate
   Standard 3 (80 ng/ml) in duplicate
   Standard 4 (160 ng/ml) in duplicate
   Standard 5 (320 ng/ml) in duplicate
   Standard 6 (640 ng/ml) in duplicate

3. Pipette 100 μL of sample (in duplicate) into pre designated wells.

4. Incubate the micro titer plate at room temperature for sixty (60 ± 2) minutes. Keep plate covered and level during incubation.

5. Following incubation, aspirate the contents of the wells.

6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with wash buffer, invert the plate then pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual buffer. Repeat 3 times for a total of four washes.

7. Pipette 100 μL of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at room temperature for ten (10 ± 2) minutes. Keep plate covered in the dark and level during incubation.

8. Wash and blot the wells as described in Steps 5/6.

9. Pipette 100 μL of TMB Substrate Solution into each well.

10. Incubate in the dark at room temperature for precisely ten (10) minutes.

11. After ten minutes, add 100 μL of Stop Solution to each well.
12. Determine the absorbance (450 nm) of the contents of each well. Calibrate the plate reader to manufacturer’s specifications.

STABILITY OF THE FINAL REACTION MIXTURE

The absorbance of the final reaction mixture can be measured up to 2 hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.

RESULTS

1. Subtract the average background value from the test values for each sample.

2. Using the results observed for the standards construct a Standard Curve. The appropriate curve fit is that of a four-parameter logistics curve. A second order polynomial (quadratic) or other curve fits may also be used.

3. Interpolate test sample values from standard curve. Correct for sera dilution factor to arrive at the Alpha 2-Macroglobulin concentration in original samples.

LIMITATION OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the information contained in the package insert instructions and with adherence to good laboratory practice.

2. Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or deionized water, and accuracy of reagent and sample pipettings, washing technique, incubation time or temperature.

3. Do not mix or substitute reagents with those from other lots or sources.